# اثر ضد تکثیر و القای آپوپتوتیک سزکوی ترپن لاکتون پارتنولید بر روی رده سلول سرطانی پستان انسان لاین MDA-MB- 231

Anti-proliferative effect and apoptotic induction of sesquiterpene lactone parthenolide in a human breast cancer cell line MDA-MB- 231

نسا جعفری'''\*، سنبل ناظری'، ساره ارجمند"، رضا بهروزی ، کبری نعلبندی'، ستار طهماسبی انفرادی ٔ

Nesa Jafari<sup>1,2,\*</sup>, Sonbol Nazeri<sup>1</sup>, Sareh Arjmand<sup>3</sup>, Reza Behroozi<sup>2</sup>, Kobra Nalbandi<sup>1</sup> and Sattar Tahmasebi Enferadi<sup>2</sup>

۱ – گروه بیوتکنولوژی، دانشکده کشاورزی، دانشگاه بوعلی سینا همدان، ایران ۲ – پژوهشکده زیست فناوری صنعت و محیط زیست، انستیتوی ملی مهندسی ژنتیک و بیوتکنولوژی، تهران ۳ – مرکز تحقیقات پروتئین، دانشگاه شهید بهشتی، جی سی، تهران، ایران

\* نو يسنده مسئول مكاتبات، پست الكترونيكي:

\*Corresponding Author, Email;<u>jafari.nesa@gmail.com</u> (تاریخ دریافت: ۱۴۰۰/۳/۳۱ - تاریخ پذیرش: ۱۴۰۰/۸/۱۶) مقاله علمي پژوهشي

مهندسی ژنتیک و ایمنی زیستی ISSN 2588-5073 چاپی ISSN 2588-5081 الکترونیکی

دوره ۱۰، شماره ۲، پاییز و زمستان ۱٤۰۰ صفحه ۱۸۹-۱۸۳

https://dorl.net/dor/20.1001.1.25885073. 1400.10.2.3.5

DOR: 20.1001.1.25885073.1400.10.2.3.5

Genetic Engineering and Biosafety Journal Volume 10, Number 2 2022

http://gebsj.ir/

https://ecc.isc.ac/showJournal/23064

چکیده

## واژههای کلیدی

اپوپتوزیس، رده سلولی MDA-MB- 231، پارتنولید، پارتنولید، RT-PCR

(Tanacetum) پارتنولید یک متابولیت ثانویه است که به طور طبیعی در گیاه بابونه کبیر parthenium و parthenium

Department of Plant Biotechnology, Faculty of Agriculture, Bu-Ali Sina University, Hamadan, Iran.

<sup>&</sup>lt;sup>2</sup> Departments of Energy and Environmental Biotechnology, National Institute of Genetic Engineering and Biotechnology, Tehran, Iran
<sup>3</sup> Protein Research Center, Shahid Beheshti University, G.C, Tehran, Iran.

# DOR: 20.1001.1.25885073.1400.10.2.14.6

# **Genetic Engineering and Biosafety Journal Volume 10, Number 2, 2022**

### **Abstract**

Parthenolide is a secondary metabolite, which naturally occurs in the feverfew plant (*Tanacetum* parthenium) and is responsible for its healing power. The potential of parthenolide in inhibition of cancer cell growth, alone or in combination with other anti-cancer therapeutics, has been studied in several laboratories. In this study, the effect of extracted parthenolide on the expression of seven pro-apoptotic genes (BID, P21, PUMA, BAX2, P53, CASP8 and BIM), all of them were influenced by NF-kB signaling pathway, in MDA-MB- 231, a breast cancer cell line was investigated. The results indicated that in response to the parthenolide treatment, all of the selected genes were induced and up-regulated significantly with different degrees. We proved that parthenolide alters the expression of a variety of genes involved in apoptosis pathway in MDA-MB- 231 cell line. In conclusion, we provided evidence that parthenolide alters the expression of a variety of genes involved in apoptosis pathway in MDA-MB- 231 breast cancer cells. The selected genes are directly or indirectly were regulated by NF-κB and it is confirmed that NF-kB is an important target of parthenolide.

**Keyword:** Apoptosis, MDA-MB- 231 cell line, Parthenolide, Real time PCR

### Introduction

Compared with synthetic analogues, plant derived natural products have relatively less side effects and renewed interest have been raised for use of natural products in pharmaceutical medications. Feverfew (Tanacetum parthenium), also known as wild chamomile, is an herb with anti-inflammatory properties and historically was used to prevent migraine and headaches, and mostly used as a fever reducer (from which the name is derived)(Ahmadi et 2018). Sesquiterpene lactone parthenolide, secondary metabolite, is the active ingredient, which is responsible for much of the healing power of feverfew and has high value in chemotaxonomy and medicine (Shahhoseini et al. 2019). Parthenolide a gemacranolide-type sesquiterpene lactone is the major constituent of European feverfew (Pooja et al. 2021). Parthenolide is also known for its exceptional anti-inflammatory and anti-cancer properties, which make it a promising candidate for further studies in drug development. Parthenolide has shown cytotoxic effects on cervical cancer (Jin et al. 2009) glioblastoma (Anderson et al. 2008) breast cancer (Nakshatri et al. 2004; Sweeney et al. 2005) leukemia (Guzman et al. 2005; Hewamana et al. 2008; Neelakantan et al. 2009) pancreatic cancer (Ramachandran et al. 2010) prostate cancer (Shanmugam et al. 2006; Sweeney et al. 2004) lung cancer, (Jin et al. 2009; Sun et al. 2020) melanomas

(Kim et al. 2007; Suvannasankha et al. 2008) and colon cancer (Zhang et al. 2004) however, it does not affect normal cells.

Anti-tumor activity of parthenolide has been shown to be strongly linked to the reactive oxygen species (ROS) generation and inhibition of transcription factor nuclear factor-kappa B (NF-κB) (Zhang et al. 2020). In combination with several common chemotherapy agents, parthenolide augments the cancer cells' response to the therapeutic agents, induces potent apoptosis and restores the cell sensitivity to the cells that developed resistance to chemotherapy.

It has been reported that the inhibition of NF-κB leads to change in the expression of apoptosis regulator target genes (Barkett and Gilmore 1999; Karam et al. 2021: Kolenko et al. 1999). In the present study, we evaluate the effect of Parthenolide on the expression of a group of pro-apoptotic target genes in MDA-MB-231 breast cancer cell line using real time PCR.

# MATERIALS AND METHODS

### Parthenolide purification

Tanacetum parthenium L.schulz Bip. Plant was collected from Hamadan province (34.7982° N, 48.5146° E) in Iran and was identified and registered by deposit number 006420 in herbarium of Tehran University. The amount of 5 g flower was isolated and ground to a fine powder in liquid nitrogen. Parthenolide Extraction was performed with 15 ml methanol: formic acid (1000:1 v/v) method of Majdi accordingly (Majdi et al. 2011). Purification of parthenolide was carried out using a preparative HPLC system equipped with a photodiode array detector and mobile phase water (A) and acetonitrile: methanol (9: 1) (B) (Avula et al. 2006)Jafari et al. 2018).

### Cell culture

MDA-MB-231 cell line was donated by Dr. Mosa Gardaneh (NIGEB, Iran). MDA-MB-231 cells were maintained in DMEM medium with 100 U.mL-1 of penicillin, 100  $\mu$ g.mL<sup>-1</sup> of streptomycin, and 10% fetal bovine serum (FBS), at 37 °C containing 5% humidified CO<sub>2</sub>. Culture medium was replaced with a fresh one every two days.

### Anti-proliferative assay

The effect of different concentrations of parthenolide (0.5, 1, 1.5, 2, 2.5, 5, 10 and 20  $\mu$ M) on the MDA-MB-231 breast cancer cell line viability was carried out by Methylthiazol Tetrazolium (MTT) for 24 hours (Jafari et al. 2018).

### Annexin-V-FLUOS assay

The annexin-V-FLUOS assay was performed to measure the percentage of apoptotic and necrotic cells under the Parthenolide treatment using Annexin-V-FLUOS staining kit (Rosche, 11988549001). MDA-MB-231 cells were treated with the minimum lethal dose (2µM) that kills approximately 50% of cells (IC50), obtained by MTT test, for 24 h. The cells were stained by FITCconjugated Annexin-V-FLUOS and PI labeling according solution to the manufacturer's recommended concentrations and time and analyzed by fluorescent microscopy. The positive cells for annexin-V and PI were considered to be apoptotic and necrotic, respectively.

### Real-time PCR

Total cell RNA was extracted using RNX-plus kit (Cinna Gen, RN7713C), after cDNA synthesis using reverted first strand cDNA synthesis kit (Thermofisher scientific, 00168871) and by using random primer, Real time PCR amplification reactions were performed using SYBR Fast qPCR kit (Roche, 03515869001) with appropriate primers (table1).

Table 1. Primers used for real time PCR

Product size	Annealing temperature	Primer sequence	Gene name	Accession numbers
256	59	F- GTCAGTGGTGGACCTGACCT R- CACCACCCTGTTGCTGTAGC	GAPDH	XR002004287.1 XM0199661648.1
362	60	F-GACAAGAATCCGACCAAATGGCAAA	BIM	LM463826.1
		R-AAAAGGATCCATGAGAAATCCTTGTGG		XM019021358.1
133	60	F-GAGGTTGGCTCTGACTGTACC	P53	XM019944223.1
		R-TCCGTCCCAGTAGATTACCAC		XM004058511.1
180	60	F-ATTAGGGACAGGAATGGAACAC	CASP8	XM002004922.1
		R-GGAGAGGATACAGCAGATGAAG		XM019838464
175	60	F-TGCCTCAGGATGCGTCCACCAA R-CCCCAGTTGAAGTTGCCGTCAG	BAX	XM0040611100 XM019978503.1
100	60	F-CCTTGCTCCGTGATGTCTTTC	BID	XM003988294.4
		R-TCCGTTCAGTCCATCCCATTT		XM016939651.1
101	60	F-GACGACCTCAACGCACAGTA	PUMA	AF354655.1
		R-AGGAGTCCCATGATGAGATTG		XM019934993.1
90	60	F-GGAGACTTCTCAGGGTCGAAAAC	P21	XM19780435.1
		R-GGGCTTCCTCTTGGAGAAGATC		XM015237492.1

A two step program was used as follows: 3 min at 95°C; 40 cycles of 10s at 95°C, 30s at 60°C which was followed by a melting curve analysis step in the Rotor-Gen 3000 Real-time DNA analysis system (Corbett, RO80902).

The relative expression ratio (RER) was measured based on the expression ratio of the target gens (Table 1) versus a reference gene (Glyceraldehyde-3-Phosphate Dehydrogenase (*GAPDH*). The relative gene expression ratio was calculated by following mathematical model.

 $\Delta C_t$  treatment =  $C_t$  (Target Gene, treatment) -  $C_t$  (GAPDH, treatment)

 $\Delta C_t$  control =  $C_t$  (Target Gene, control) –  $C_t$  (GAPDH, control)

RER =  $2^{-\Delta\Delta C}_{t}$  (Livak and Schmittgen 2001).

### **Statistical Analysis**

The Statistical Package for Social Sciences (SPSS) software was used to Statistical analysis. T-test was performed for finding significant differences in gene expression values. Differences with P values < 0.01 were considered as significant. Orthogonal comparisons were used to evaluate the significant difference between the means of treatments compared to the control.

# **RESULTS AND DISCUSSION**

Studies have shown that in Feverfew (*Tanacetum parthenium*), parthenolide accumulates in flower (Majdi et al. 2011). Accordingly, we extracted parthenolide from flower head and then purified with HPLC-Preparative (Figure 1-A).

Programmed cell death or apoptosis is one of the major animal cell death types which is characterized by many changes in morphological and molecular level, including cell shrinkage, nuclear condensation, plasma membrane blebbing, and differentially expression of many genes related in the apoptosis related pathways (Zhang et al. 2009).

Herein, the MTT assay results, confirmed that the purified parthenolide behaves as an apoptotic inducer for MDA-MB-231 cells (IC<sub>50</sub> value= 1.5  $\mu$ M) (Figure 1-B). This result verified by the Annexin-V-FLUOS staining (Figure 2).

Describing the molecular mechanisms of this apoptotic induction may be applicable to improve the chemotherapeutic strategies for breast cancer.

Several studies have recognized that induction of apoptosis by parthenolide is associated with inhibition of NF- $\kappa$ B (Pozarowski et al. 2003; Saadane et al. 2007; Yip-Schneider et al. 2005) and increased reactive oxygen species (D'Anneo et al. 2013). For instance, Liu and et al showed that parthenolide can decrease the protein expression level of anti-apoptotic BcL2 and increase protein expression of proapoptotic BAX in pancreatic cancer cells (Liu et al. 2010). Both BcL2 and BAX are regulated by NF- $\kappa$ B.

For more clarification of the molecular mechanism of parthenolide-induced apoptosis, in this study seven pro-apoptotic genes were selected. Expression of these genes in the MDA-MB-231 cells to be studied when was treated by parthenolide and compared with control cells. All of the selected genes, including *BID*, *P21*, *PUMA*, *BAX2*, *P53*, *CASP8* and *BIM*, are affected by NF-κB activation. The results of real time PCR indicated the up-regulation of all genes in response to parthenolide treatment (Figure 3).

However, the increased amount is very different. Differences between the genes expression in before and after treatment were calculated according to Livak mathematical model. Changes were mostly observed in BID gene expression with almost 25-fold increase in the expression compared with control cells. BID, an acronym for BH3-interacting domain death agonist, is a pro-apoptotic BCL-2 protein family. After cleavage with CASP8, activated BID translocates from cytosol to mitochondria and induces apoptosis by promoting BAX activation and mitochondrial outer membrane permeabilization, which results in the release of cytochrome c and other pro-apoptotic factors. It is shown that blockage of NF-κB signaling prevents the cleavage of BID (Danthi et al. 2010).

Studies have proven that targeting NF-kB through parthenolide could indirectly influence the proapoptotic activity of *P53* (Kim et al. 2009). Parthenolide can regulate *P53* activity by increasing MDM<sub>2</sub> ubiquitination (Gopal et al. 2009). *P21*, is a cell cycle inhibitor with a major role in growth arrest, and is tightly regulated by *P53*. The up-regulation of *P21* in *P53*-independent manner has also been described in response to certain stresses including DNA damage (Hosseini et al. 2017; Macleod et al. 1995).

Here, a five-fold increase was observed in P21 expression while P53 expression level compared with control cells, has changed slightly ( $\sim 1.2$ -fold increase). Hence, it seems that Parthenolide, in this study, most likely up regulates the P21 independently of P53.

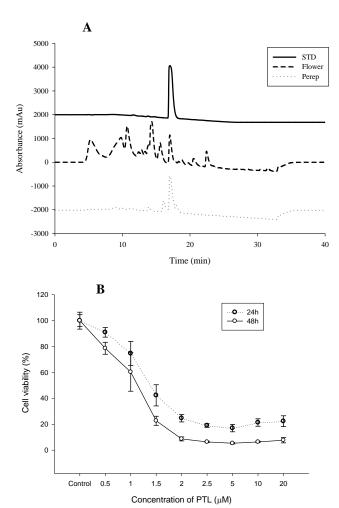


Fig 1. (A) HPLC chromatogram of parthenolide standard (STD), flower extract of feverfew (Flower) and purified parthenolide from feverfew by preparative HPLC. (B) MTT assay results. Viability of MDA-MB-231 cells treated with Parthenolide decreased in a dose dependent manner and the IC<sub>50</sub> value for MDA-MB-231 was calculated 1.5  $\mu$ M. Data are shown as means  $\pm$  SD (n=3).

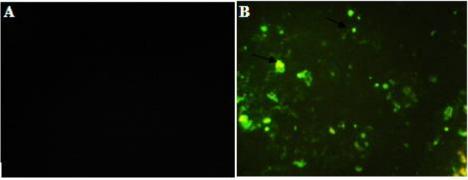


Fig 2. Annexin-V-FLUOS staining of MDA-MB-231 cells. (A)Untreated cells, and (B) treated cells with  $2\mu M$  parthenolide for 24h. Early apoptotic cells were stained with green fluorescent Annexin V. No necrotic cells were detected.

Fig 3. Real time expression analysis of *BID*, *P21*, *PUMA*, *CASP8*, *BAX*, *P53*, and *BIM* genes. The relative expression ratio (R) was measured based on the expression ratio of the target gens versus a reference gene (Glyceraldehyde-3- Phosphate Dehydrogenase (GAPDH) in control and treatment cells. The results showed that the expression of all selected genes in MDA-MB-231 cell treated with Parthenolide induced and up-regulated. Data are shown as means  $\pm$  SD (n=3).

In conclusion, we provided evidence that parthenolide alters the expression of a variety of genes in treatment cells compared with control cells, involved in apoptosis pathway in MDA-MB- 231 breast cancer cells. The selected genes are directly or indirectly regulated by NF- $\kappa$ B and it is confirmed that NF- $\kappa$ B is an important target of parthenolide.

### Acknowledgements

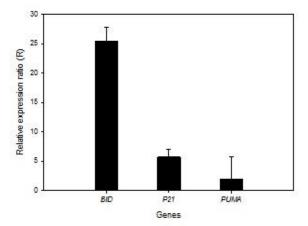
This work was supported by the National Institute of Genetic Engineering and Biotechnology of IRAN.

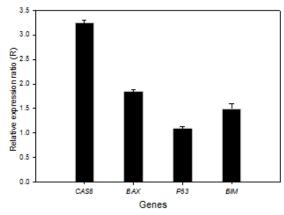
### **Conflict of Interest Statement**

The authors declare that no conflict of interest regarding the publication of this paper

# REFERENCES

- Ahmadi SZ, Ghorbanpour M, Hadian J, Salehi-Arjmand H (2018) Impact of foliar spray of spherical nano-carbon and salicylic acid on physiological traits and parthenolide content in two Feverfew cultivars (Tanacetum parthenium Linn. cv. Pharmasaat and Jelitto). Journal of Medicinal Plants 17
- Anderson JC, McFarland BC, Gladson CL (2008) New molecular targets in angiogenic vessels of glioblastoma tumours. Expert reviews in molecular medicine 10: e23
- Avula B, Navarrete A, Joshi V, Khan I (2006) Quantification of parthenolide in Tanacetum species by LC-UV/LC-MS and microscopic comparison of Mexican/US feverfew samples. Die Pharmazie-An International Journal of Pharmaceutical Sciences 61: 590-594
- Barkett M, Gilmore TD (1999) Control of apoptosis by Rel/NFkappaB transcription factors. Oncogene 18: 6910-6924
- D'Anneo A, Carlisi D, Lauricella M, Puleio R, Martinez R, Di Bella S, Di Marco P, Emanuele S, Di Fiore R, Guercio A (2013) Parthenolide generates reactive oxygen species and autophagy in MDA-MB231 cells. A soluble parthenolide analogue inhibits tumour growth and metastasis in a xenograft model of breast cancer. Cell death & disease 4: e891
- Danthi P, Pruijssers AJ, Berger AK, Holm GH, Zinkel SS, Dermody TS (2010) Bid regulates the pathogenesis of neurotropic reovirus. PLoS Pathog 6: e1000980





Gopal YV, Chanchorn E, Van Dyke MW (2009) Parthenolide promotes the ubiquitination of MDM2 and activates p53 cellular functions. Molecular cancer therapeutics 8: 552-562

- Guzman ML, Rossi RM, Karnischky L, Li X, Peterson DR, Howard DS, Jordan CT (2005) The sesquiterpene lactone parthenolide induces apoptosis of human acute myelogenous leukemia stem and progenitor cells. Blood 105: 4163-4169
- Hewamana S, Alghazal S, Lin TT, Clement M, Jenkins C, Guzman ML, Jordan CT, Neelakantan S, Crooks PA, Burnett AK (2008) The NF-κB subunit Rel A is associated with in vitro survival and clinical disease progression in chronic lymphocytic leukemia and represents a promising therapeutic target. Blood 111: 4681-4689
- Hosseini FS, Falahati-pour SK, Hajizadeh MR, Khoshdel A, Mirzaei MR, Ahmadirad H, Behroozi R, Jafari N, Mahmoodi M (2017) Persian shallot, Allium hirtifolium Boiss, induced apoptosis in human hepatocellular carcinoma cells. Cytotechnology: 1-13
- Jin X, Qiu L, Zhang D, Zhang M, Wang Z, Guo Z, Deng C, Guo C (2009) Chemosensitization in non-small cell lung cancer cells by IKK inhibitor occurs via NF-κB and mitochondrial cytochrome c cascade. Journal of cellular and molecular medicine 13: 4596-4607
- Karam L, Abou Staiteieh S, Chaaban R, Hayar B, Ismail B, Neipel F, Darwiche N, Abou Merhi R (2021) Anticancer activities of parthenolide in primary effusion lymphoma preclinical models. Molecular Carcinogenesis
- Kim DH, Bae J, Lee JW, Kim SY, Kim YH, Bae JY, Yi JK, Yu MH, Noh DY, Lee C (2009) Proteomic analysis of breast cancer tissue reveals upregulation of actin-remodeling

- proteins and its relevance to cancer invasiveness. PROTEOMICS-Clinical Applications 3: 30-40
- Kim KW, Cho ML, Kim HR, Ju JH, Park MK, Oh HJ, Kim JS, Park SH, Lee SH, Kim HY (2007) Up-regulation of stromal cell–derived factor 1 (CXCL12) production in rheumatoid synovial fibroblasts through interactions with T lymphocytes: Role of interleukin-17 and CD40L–CD40 interaction. Arthritis & Rheumatism 56: 1076-1086
- Kolenko V, Bloom T, Rayman P, Bukowski R, Hsi E, Finke J (1999) Inhibition of NF-kappa B activity in human T lymphocytes induces caspase-dependent apoptosis without detectable activation of caspase-1 and -3. Journal of immunology 163: 590-598
- Liu J-W, Cai M-X, Xin Y, Wu Q-S, Ma J, Yang P, Xie H-Y, Huang D-S (2010) Parthenolide induces proliferation inhibition and apoptosis of pancreatic cancer cells in vitro. Journal of Experimental & Clinical Cancer Research 29: 1
- Livak KJ, Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2–ΔΔCT method. methods 25: 402-408
- Macleod KF, Sherry N, Hannon G, Beach D, Tokino T, Kinzler K, Vogelstein B, Jacks T (1995) p53-dependent and independent expression of p21 during cell growth, differentiation, and DNA damage. Genes & development 9: 935-944
- Majdi M, Liu Q, Karimzadeh G, Malboobi MA, Beekwilder J, Cankar K, de Vos R, Todorović S, Simonović A, Bouwmeester H (2011) Biosynthesis and localization of parthenolide in glandular trichomes of feverfew (Tanacetum parthenium L. Schulz Bip.). Phytochemistry 72: 1739-1750
- Nakshatri H, Rice SE, Bhat-Nakshatri P (2004) Antitumor agent parthenolide reverses resistance of breast cancer cells to tumor necrosis factor-related apoptosis-inducing ligand through sustained activation of c-Jun N-terminal kinase. Oncogene 23: 7330-7344
- Neelakantan S, Nasim S, Guzman ML, Jordan CT, Crooks PA (2009) Aminoparthenolides as novel anti-leukemic agents: Discovery of the NF-κB inhibitor, DMAPT (LC-1). Bioorganic & medicinal chemistry letters 19: 4346-4349
- Pooja S, Shetty P, Kumari N, Shetty K (2021) Radioprotective and antioxidant potential of Tanacetum parthenium extract and synthetic parthenolide in Swiss albino mice exposed to electron beam irradiation. International Journal of Radiation Research 19: 145-154
- Pozarowski P, Huang X, Halicka D, Lee B, Johnson G, Darzynkiewicz Z (2003) Interactions of fluorochrome-labeled caspase inhibitors with apoptotic cells: A caution in data interpretation. Cytometry Part A 55: 50-60
- Ramachandran C, Resek AP, Escalon E, Aviram A, Melnick SJ (2010) Potentiation of gemcitabine by Turmeric Force™ in pancreatic cancer cell lines. Oncology reports 23: 1529-1535
- Saadane A, Masters S, DiDonato J, Li J, Berger M (2007) Parthenolide inhibits IκB kinase, NF-κB activation, and inflammatory response in cystic fibrosis cells and mice.

- American journal of respiratory cell and molecular biology 36: 728-736
- Shahhoseini R, Azizi M, Asili J, Moshtaghi N, Samiei L (2019)
  Comprehensive Assessment of Phytochemical Potential of
  Tanacetum parthenium (L.): Phenolic Compounds,
  Antioxidant Activity, Essential Oil and Parthenolide. Journal
  of Essential Oil Bearing Plants 22: 614-629
- Shanmugam R, Jayaprakasan V, Gokmen-Polar Y, Kelich S, Miller KD, Yip-Schneider M, Cheng L, Bhat-Nakshatri P, Sledge GW, Nakshatri H (2006) Restoring chemotherapy and hormone therapy sensitivity by parthenolide in a xenograft hormone refractory prostate cancer model. The Prostate 66: 1498-1511
- Sun L, Yuan W, Wen G, Yu B, Xu F, Gan X, Tang J, Zeng Q, Zhu L, Chen C (2020) Parthenolide inhibits human lung cancer cell growth by modulating the IGF-1R/PI3K/Akt signaling pathway. Oncology Reports 44: 1184-1193
- Suvannasankha A, Crean CD, Shanmugam R, Farag SS, Abonour R, Boswell HS, Nakshatri H (2008) Antimyeloma effects of a sesquiterpene lactone parthenolide. Clinical Cancer Research 14: 1814-1822
- Sweeney C, Li L, Shanmugam R, Bhat-Nakshatri P, Jayaprakasan V, Baldridge LA, Gardner T, Smith M, Nakshatri H, Cheng L (2004) Nuclear factor-kB is constitutively activated in prostate cancer in vitro and is overexpressed in prostatic intraepithelial neoplasia and adenocarcinoma of the prostate. Clinical Cancer Research 10: 5501-5507
- Sweeney CJ, Mehrotra S, Sadaria MR, Kumar S, Shortle NH, Roman Y, Sheridan C, Campbell RA, Murry DJ, Badve S (2005) The sesquiterpene lactone parthenolide in combination with docetaxel reduces metastasis and improves survival in a xenograft model of breast cancer. Molecular cancer therapeutics 4: 1004-1012
- Yip-Schneider MT, Nakshatri H, Sweeney CJ, Marshall MS, Wiebke EA, Schmidt CM (2005) Parthenolide and sulindac cooperate to mediate growth suppression and inhibit the nuclear factor-κB pathway in pancreatic carcinoma cells. Molecular cancer therapeutics 4: 587-594
- Zhang D, Qiu L, Jin X, Guo Z, Guo C (2009) Nuclear Factor-κB Inhibition by Parthenolide Potentiates the Efficacy of Taxol in Non–Small Cell Lung Cancer In vitro and In vivo. Molecular Cancer Research 7: 1139-1149
- Zhang S, Ong C-N, Shen H-M (2004) Critical roles of intracellular thiols and calcium in parthenolide-induced apoptosis in human colorectal cancer cells. Cancer letters 208: 143-153
- Zhang Y, Huang Q, Chen Y, Peng X, Wang Y, Li S, Wu J, Luo C, Gong W, Yin B (2020) Parthenolide, an NF-κB inhibitor, alleviates peritoneal fibrosis by suppressing the TGF-β/Smad pathway. International immunopharmacology 78: 106064